



A Combinatorics Approach to Lipid A Structure Identification

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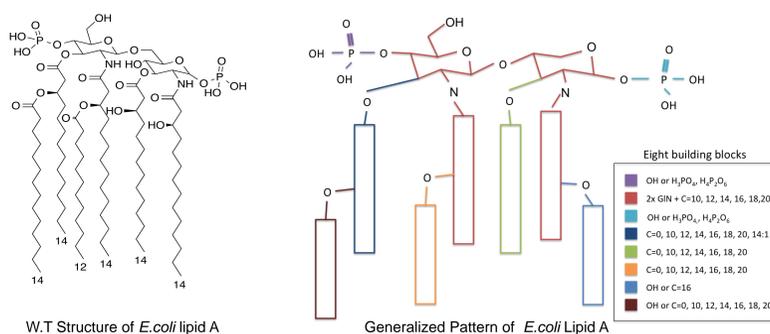
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Introduction

Lipid A, the major component of Gram-negative bacterial outer membrane, is also known as endotoxin and is recognized by host immune system. Recently, we reported on the hierarchical tandem mass spectrometry (HiTMS) algorithm (Ting et al. *JASMS* 2011) for the assignment of lipid A structures, the hydrophobic anchor of lipopolysaccharide. This approach required acquisition of exhaustive tandem MSⁿ data on multiple precursor ions and subsequent fragment ions. However, due to sample availability, it is not an appropriate technique for clinically relevant samples. Therefore, we are developing a combinatorics approach to solve these unique glycolipid structures. This new approach results in a probabilistic match to a structure that requires only tandem mass spectra (MS²).

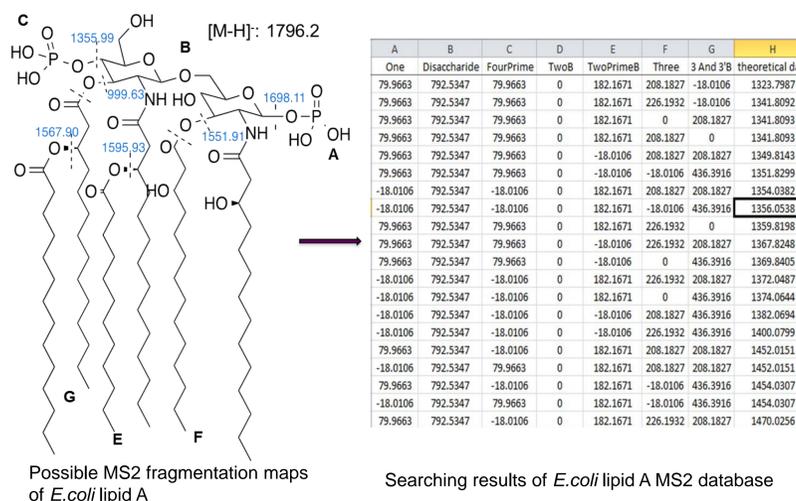
In this study, theoretical structures for *E. coli* lipid A were computed using a combinatorics algorithm that considered the heterogeneity present in the fatty acids (number of carbon chain length and position variety) and the phosphorylation pattern on the disaccharide backbone. Precursor ion masses detected from a lipid A structure were compared against the MS¹ theoretical database we constructed and the top n selected based on a match to the available molecular masses. A hypothetical structure is further elucidated by matching the tandem mass spectra against a secondary MS/MS database consisting of all the possible fragmentation patterns based on the combinatorics algorithm.

MS¹ Theoretical Database Construction



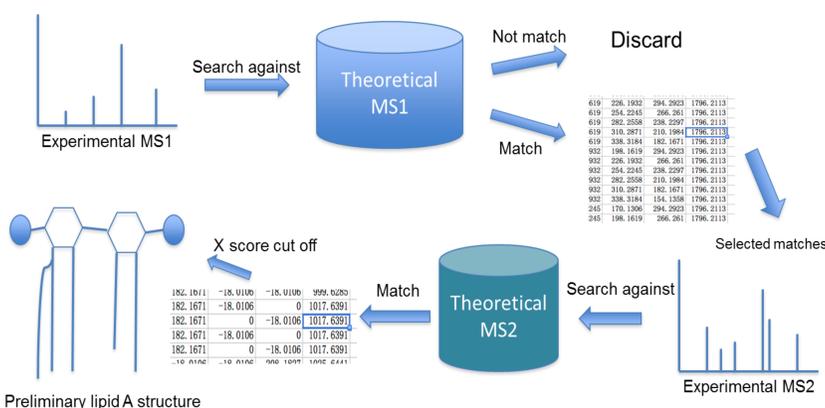
Based on manual interpretation of lipid A precursor ion (MS¹) data and fragmentation rules (Ting et al. *JASMS* 2011), eight components (fatty acid, phosphate pattern, disaccharide et al.) of potential theoretical masses derived from the *E. coli* lipid A structure were calculated. Using our combinatorial algorithm, each component is considered as a module to construct a hypothetical lipid A structure.

MS² Theoretical Database Construction



For MS², possible fragment patterns determined from literatures reported tandem mass spectrometry data (John P. O'Brien, et. al, *Ana. Chem.* 2014, Chang-soo Lee et al., *JASMS*, 2004). Based on these, *E. coli* lipid A fragmented map was drawn and all possible fragment ion masses were calculated.

Proposed Flowchart of Combinatorics Algorithm



All possible component configurations were calculated to generate theoretical *E. coli* lipid A MS¹ data (~300,000 possible combinations) and incorporated into the program implemented in Python 2.7.6. Acquired MS¹/MS² experimental data were manually verified by searching against lipid A MS¹/MS² theoretical database.

Conclusions

- All possible exact masses for precursor ion were calculated based on the generalized *E. coli* lipid A structure resulting in a total of 296,352 possible combinations and incorporated into a MS¹ lipid A database.
- All possible exact masses for fragment ions were calculated based on the proposed MS² fragmentation map and manually verified by using previously published diagnostic fragment ions. Of these 12 featured fragment ions, all were matched in MS² theoretical database which contained 128 possible ions for m/z of 1796 [M-H]⁻ precursor ion.
- Preliminary results of our combinatorics algorithm for Lipid A structure identification are promising. While the initial work was focused on elucidating the lipid A structure of *E. coli*, we are working on testing the approach against a variety of bacterial species lipid A structures that show heterogeneity in both the length and number of fatty acids as well as modification of the phosphate group attached to the diglucosamine backbone.

References

- Ting, Ying S., et al. *J. Am. Soc. Mass Spectrom* **22** (2011): 856-866.
- Chang-soo Lee et al., *J. Am. Soc. Mass Spectrom.*, **39** (2004): 514-525.
- O'Brien, John Patrick, et al. *Anal. Chem.* **86** (2014) : 2138-2145
- Needham, Brittany D., and M. Stephen Trent. *Nat. Rev. Micro.* **11** (2013): 467-481.

Acknowledgements

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